over time, or if the microplastics would be easily passed with fecal matter. This is an area we would suggest for future research. If these plastic fibers are unable to be easily passed and accrue over time, they could lead to reduced nutritional uptake or impaction. Furthermore, there remains the potential for chemical leaching as these secondary microplastics are further broken down as they pass through the salamander's digestive tract. Either way, there remains the potential for these ingested plastics to have long-lasting negative effects on salamander health. Given that increasing studies have found that microplastics pose a significant threat to wildlife health (Sarkar et al. 2023. Int. J. Environ. Res. Public Health 20:1745) and are found across most ecosystems (e.g., from the deep sea to the high Arctic, but increasing in concentration in urban areas), the potential for this widespread form of pollution to negatively impact amphibians is notable and merits further investigation.

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***TARICHA TOROSA* (California Newt). DIET.** *Taricha torosa* is found along the Pacific Coast of California ranging from Mendocino County to San Diego County (McGinnis and Stebbins 2018. Field Guide to Reptiles and Amphibians of California. Houghton Mifflin, Harcourt Publishing, Boston, Massachusetts. 560 pp.). *Taricha torosa* is relatively common north of Monterey County but is declining rapidly from south of Monterey County to northern San Diego County (Thomson et al. 2016. California Amphibian and Reptile Species of Special Concern. University of California Press, Berkeley, California. 390 pp.). Ritter (1897. Proc. Calif. Acad. Sci. 1:73–114) described *T. torosa* larvae feeding on decomposing organic matter and suggested the possibility of cannibalism. Storer (1925. A Synopsis of the Amphibia of California. University of California Press, Berkeley, California. 343 pp.) reported captive *T. torosa* feeding on mosquito larvae. Additional feeding behavior was also looked at by Marchetti and Hayes (2020. West. N. Am. Nat. 80:165–174) and Marchetti et al. (2022. West. N. Am. Nat. 82:77–85), which showed that larvae tend to have an ontogenetic change in foraging behavior as they grow—larger larvae eat larger food items. Marchetti et al. (2022, *op. cit.*) found larval stomach contents to be largely comprised of arthropods and gastropods, with chironomids (non-biting midges) being the most abundant prey item, while gastropods represented the second most common prey item. Thus, *T. torosa* larvae appear to be primarily predators of small aquatic invertebrates. Here, we report two separate observations that not only support the hypothesis of Ritter (1897, *op. cit.*) but indicate larval *T. torosa* may be capable of preying on other amphibian larvae.

In September 2023, while conducting a translocation study for *Rana draytonii* (California Red-legged Frog) in an artificial stock pond, we observed *T. torosa* larvae (ca. 45 mm total length) opportunistically scavenging on a dead *Pseudacris regilla* larva (Pacific Chorus Frog) (Gosner Stage 43–45) at a pond in Napa County, California, USA (38.45410°N, 122.16979°W; WGS 84; Fig. 1). As we continued surveying the pond, we also observed a similar sized *T. torosa* larva feeding on the rear leg of a live *P. regilla* larva (Gosner Stage 41–42) and dragging it down into the mid-water column of the pond. We did not observe the outcome.

*Taricha* consumption of anuran eggs has been previously documented. Westphal et al. (2021. Herpetol. Rev. 52:820) reported *T. granulosa* larvae consuming anuran (i.e., *R. draytonii*) eggs and

FIG. 1. A dead *Pseudacris regilla* larva being opportunistically consumed by two larval *Taricha torosa* in Napa County, California, USA.

Rathbun (1998. Herpetol. Rev. 29:165) also reported adult *Taricha* feeding on anuran eggs. Our observations of *T. torosa* larvae consuming *P. regilla* larvae are noteworthy as this is the first report of *Taricha* consumption of anuran larvae.

González-Mollinedo and Mármol-Kattán (2019. Herpetol. Rev. 50:762) reported that larval scavenging could increase the transmission risk of chytridiomycosis to the scavenging individual. Alvarez and Wilcox (2021. Herpetol. Rev. 52:821) suggested that there should be consideration when research is conducted on populations of larvae known to scavenge on other larvae that are known to be infected by chytridiomycosis, ranavirus, endoparasites or other pathogens. Our observation suggests that larval *T. torosa* do predate and engage in necrophagia with sympatric larval amphibians as they reach a size where doing so promotes growth.

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