

Acknowledgements—To the Secretaría de Ciencia, Humanidades, Tecnología e Innovación (SECIHTI), for posdoctoral scholarship to GS-V (CVU: 693584).

Literature Cited

Bauer, A.M. 1997. Peritoneal pigmentation and generic allocation in the Chameleonidae. *African Journal of Herpetology* 46:117-123.

Franco-Martínez, J.R.P., A. González-Huerta, D.J. Pérez-López, and M. González-Ronquillo. 2015. Phenotypic characterization of hybrids and varieties of forage maize in High Valley State of Mexico, Mexico. *Revista Mexicana de Ciencias Agrícolas* 8:1915-1927.

Goldberg, J., B.S.L. Valverde, and L. Franco-Belussi. 2020. Testicular melanization in anuran species: ontogeny and sexual maturity. *Amphibia-Reptilia* 4:75-86.

Gómez-Benítez, A., E.A. Reyes-Velázquez, J. Rheubert, and O. Hernández-Gallegos. 2025. Ontogenetic allometry and non-size-dependent variation of head shape in the Mexican Dusky Rattlesnake. *Revista Mexicana de Biodiversidad* 96:1-10.

Gribbins, K.M., and J.L. Rheubert. 2014. The architecture of the testis, spermatogenesis, and mature spermatozoa. Pages 340-424 in: J.L. Rheubert, D.S. Siegel, and S.E. Trauth (Eds.). *Reproductive biology and phylogeny of lizards and tuatara*. CRC Press, Boca Raton, FL.

Guillette, L.J., J. Weigel, and G. Flater. 1983. Unilateral testicular pigmentation in the Mexican lizard *Sceloporus variabilis*. *Copeia* 1983:155-161.

Klaver, C. 1979. A review of *Brookesia* systematics with special reference to lung-morphology (Reptilia, Sauria, Chamaeleonidae). *Bonner Zoologische Beiträge* 30:162-175.

Lee, J.L., A.H. Miller, G.R. Zug, and D.G. Mulcahy. 2019. The discovery of Rock Geckos *Cnemaspis* Strauch, 1887 (Squamata: Gekkonidae) in the Tanintharyi Region, Myanmar with the description of two new species. *Zootaxa* 4661:40-64.

Meunier, L., G. Sorci, C. Silva Vieira, Y. Hingrat, M. Saint Jalme, and J. Torres Carreira. 2024. Sperm morphology, and macro and microscopic description of male and female reproductive tract in the North African houbara bustard (*Chlamydotis undulata*). *Theriogenology Wild* 4:1-22.

Neaves, W.B. 1976. Structural characterization and rapid manual isolation of a reptilian testicular tunic rich in Leydig cells. *The Anatomical Record* 186:553-564.

Parsley, L.M., E. Wapstra, and S.M. Jones. 2015. Atrazine disrupts gonadal development in a live-bearing lizard. *Endocrine Disruptors* 3:1-10.

Pewhom, A., and N. Srakaew. 2018. Microanatomy of the testes and testicular ducts of the butterfly lizard, *Leiolepis ocellata* Peters, 1971 (Reptilia: Squamata: Agamidae) during the active reproductive period. *Acta Zoologica* 101:1-18.

Porter, W.P. 1967. Solar radiation through the living body walls of vertebrates with emphasis on desert reptiles. *Ecological Monographs* 37:273-296.

Sanz-Ochotorena, A., M.L. Segura-Valdés, Y. Rodríguez-Gómez, R. Lara-Martínez, and L.F. Jiménez-García. 2011. Pigmentos en los testículos de cinco anfibios endémicos de Cuba (*Eleutherodactylus turquinensis*, *E. cuneatus*, *E. glamyrus*, *Bufo longinasus longinasus* y *B. longinasus cajalbanensis*). *Revista Especializada en Ciencias Químico-Biológicas* 14:30-37.

NATURAL HISTORY NOTE

Biofluorescence from the Skin Toxin in the California Red-legged Frog in Central California

Jeff A. Alvarez, The Wildlife Project, P.O. Box 188888, Sacramento, CA; jeff@thewildlifeproject.com

Reports of biofluorescence in wildlife have included numerous species of mammals, birds, reptiles, amphibians, and invertebrates (Lawrence 1954, Babu et al. 2002, Honkavaara et al. 2002, Maxwell and Johnson 2000, McGraw and Nogare 2004, Lagorio et al. 2015). Authors have reported that biofluorescence typically occurs when tissues absorb electromagnetic radiation (e.g., ultraviolet light) at a relatively high wavelength, which are then re-emitted at a lower wavelength, with visually detectable fluorescing light from specific tissues. Biofluorescence has been shown in a number of amphibian (Deschepper et al. 2018, Whitcher 2020) and reptile species (Gruber and Sparks 2015, Seiko

2019, Fuentes Magallón et al. 2021), typically under ultraviolet light (UV) excitation (Lamb and Davis 2020). Several authors reported biofluorescence from the bones of frogs and lizards (Prötzler et al. 2018, Goutte et al. 2019, Alvarez et al. 2025a), but the majority of reports of biofluorescence has been restricted to the skin (Taboda et al. 2017, Gray 2019, Kong et al. 2023, Alvarez et al. 2025b), or reflectance from the eyes (Alvarez et al. 2022, Alvarez and Perpignani 2024).

A wide variety of toxins have been found in the skin on frogs and toads. They are thought to be defensive but need to be concentrated in large quantities to be effective (Duellman and Trueb 1994). California anurans appear



Fig. 1. A California Red-legged Frog placed in a bucket of pond water, within 10 seconds of PIT tag placement, illuminated with an ultra-violet light showing the skin exudate biofluorescence. Photo by Jeff A. Alvarez.

to have a skin toxin that may have a unique odor that is detectable by humans. For example, the Arroyo Toad (*Anaxyrus californica*) can smell like raw peanuts (*Achris hypogaea*), the Foothill Yellow-legged Frog (*Rana boylii*) like the plant poison hemlock (*Conium maculatum*), and the California Red-legged Frog (*R. draytonii*) like burning rubber or plastic (pers. obs.). In the case of the California Red-legged Frog, I report that the skin toxin also appears to have the ability to be biofluorescent when exposed to ultraviolet light.

During a long-term study of California Red-legged Frogs that were the subjects of translocation from Sonoma County to Napa County, California, we collected adult and post-metamorphic (young of the year) frogs for processing. Each frog was weighed, measured and a Passive Integrated Transducer (PIT) tag was inserted under the dorsal skin surface for later identification. PIT tags were placed by gathering 1-2 mm of loose skin on the dorsal surface, approximately at the shoulder of each individual. The loose skin was then incised with a small pair of fine scissors such that a 2 mm opening was created into which a PIT tag could be inserted with canulated forceps. The tag was then manually manipulated posteriorly so that the PIT tag rested posterior to the sacral hump. Each frog was then placed in a bucket of pond water and allowed to recover for approximately 30 minutes.

When frogs were placed in buckets and exposed to ultra violet light—a 365 nm ultraviolet (UV) light (Convoy C8 + 365nm UV LED Flashlight with Patented Glass Filter) for 5 to 10 seconds I noted the freshly PIT tagged frogs, which showed no signs of toxic exudate and no indication of blood under white light, showed clear indications of biofluorescence at the wound site for the



Fig. 2. The same California Red-legged Frog 30 seconds following placement of a PIT tag showing little to no biofluorescence under an ultra-violet light. Photo by Jeff A. Alvarez.

first 10-15 seconds (Fig. 1). This reflective exudate slowly subsided and became less detectable by approximately 30 seconds (Fig. 2), and undetectable thereafter. We also noted the adults, when initially captured by hand, and exuding the skin toxin to the extent that it could easily be detected olfactorily, also emitted a biofluorescence in areas that appeared to be toxic exudate.

Many authors have reported biofluorescence from the skin of amphibians, while under ultraviolet light (Taboada et al. 2017, Gray 2019, Lamb and Davis 2020), but the ecological role of fluorescence is the subject of much consideration (Honkavaara et al. 2002, Lagorio et al. 2015, Taboada et al. 2017). Several authors have reported that a range of species may be using this type of biofluorescence as a means of interspecific communication, and even interaction among conspecifics (Lim 2007, Sparks et al. 2014, Prötz et al. 2018). It is possible that the California Red-legged Frog, and other species of California anuran, use this type of biofluorescence as a type of aposematic communication to predators that molest the animals, although that remains limited to speculation.

Acknowledgements—The Land Trust of Napa County granted access to the site where California Red-legged Frogs were PIT tagged. Frog handling and tagging was permitted under California Department of Fish and Wildlife policies: Department Bulletin 2017-04 (Captive Propagation of Fish, Wildlife and Plants for Conservation Purposes) and 2017-05 (Policy and Procedures for Conservation Translocations of Animals and Plants). I also wish to thank Nina Jackson for her assistance in capturing and PIT tagging California Red-legged Frogs.

It is possible that the California Red-legged Frog, and other species of California anuran, use this type of biofluorescence as a type of aposematic communication to predators that molest the animals, although that remains limited to speculation.

Literature Cited

Alvarez, J.A., P. Lewis-Deweese, and J.T. Wilcox. 2022. Ocular biofluorescence due to ultra-violet excitation in California red-legged and foothill yellow-legged frogs, in Central California, USA. *Sonoran Herpetologist* 35:151-153.

Alvarez, J.A., C. Noriega, J.H. Valdez-Villavicencio, and D.F. Perez-Canas. 2025a. Biofluorescence in the skeletal system of the granite night lizard (*Xantusia henshawi*) in central California. *Sonoran Herpetologist* 38:96-98.

Alvarez, J.A., I. Parr, E. Gutberlet, and N. Kleponis. 2025b. Biofluorescence in adult western spadefoot (*Spea hammondii*) in central California. *Sonoran Herpetologist* 38:88-90.

Alvarez, J.A., and R. Perpignani. 2024. *Lithobates catesbeianus* (American Bullfrog) Ultra-violet reflectivity. *Herpetological Review* 55:74-75.

Babu, B.G., and M. Kannan. 2002. Lightning bugs. *Resonance* 7:49-55.

Deschepper, P., B. Jonckheere, and J. Matthys. 2018. A light in the dark: the discovery of another fluorescent frog in the Costa Rican rainforests. *Wilderness & Environmental Medicine* 29:421-422.

Duellman, W.E., and L. Trueb. 1994. Biology of amphibians. Johns Hopkins University Press, Baltimore, MD.

Fuentes Magallón, R., et al. 2021. First record of fluorescence in Colombian long-tailed snakes, (*Enuliophis sclateri*) (Squamata: Dipsadidae), from Panama. *Reptiles & Amphibians* 28:442-443.

Goutte, S., et al. 2019. Intense bone fluorescence reveals hidden patterns in pumpkin toadlets. *Scientific Reports* 29:5388.

Gray, R.J. 2019. Biofluorescent lateral patterning on the mossy bushfrog (*Philautus macroscelis*): the first report of biofluorescence in a rhacophorid frog. *Herpetology Notes* 12:363-364.

Gruber, D.F., and J.S. Sparks. 2015. First observation of fluorescence in marine turtles. *American Museum Novitates* 3845:1-8.

Honkavaara, J., et al. 2002. Ultraviolet vision and foraging in terrestrial vertebrates. *Oikos* 98:504-510.

Kong, B., K. Preston, M. Ishimatsu, and E. Adelsheim. 2023. Biofluorescence in the California tiger salamander, *Ambystoma californiense* (Amphibia: Ambystomatidae). *Herpetology Notes* 16:161-163.

Lagorio, M.G., G.B. Cordon, and A. Iriel. 2015. Reviewing the relevance of fluorescence in biological systems. *Photochemical & Photobiological Sciences* 14:1538-1559.

Lamb, J.Y., and M.P. Davis. 2020. Salamanders and other amphibians are aglow with biofluorescence. *Scientific Reports* 10:e2821.

Lawrence, R.F. 1954. Fluorescence in arthropoda. *Journal of the Entomological Society of South Africa* 17:167.

Lim, M.L.M., M.F. Land, and D. Li. 2007. Sex-specific UV and fluorescence signals in jumping spiders. *Science* 315:481.

Maxwell, K., and G.N. Johnson. 2000. Chlorophyll fluorescence — a practical guide. *Journal of Experimental Botany* 51:659-668.

McGraw, K.J., and M.C. Nogare. 2004. Carotenoid pigments and the selectivity of psittacofulvin-based coloration systems in parrots. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* 138:229-233.

Prötzel, D., et al. 2018. Widespread bone-based fluorescence in chameleons. *Scientific Reports* 8:1-9.

Seiko, T., and Y. Terai. 2019. Fluorescence emission in a marine snake. *Galaxea* 21:7-8.

Sparks, J.S., et al. 2014. The covert world of fish biofluorescence: a phylogenetically widespread and phenotypically variable phenomenon. *PLoS One* 9:e83259.

Taboada, C., et al. 2017. Fluorescent frogs: A herpetological perspective. *South American Journal of Herpetology* 12:1-13.

Whitcher, C. 2020. New accounts of biofluorescence in several anuran genera (Hylidae, Microhylidae, Ranidae, Leptodactylidae) with comments on intraspecific variation. *Herpetology Notes* 13:443-447.

NATURAL HISTORY NOTE

Late Fall Activity in the Gray Treefrog (*Hyla versicolor*) in Northwestern and Central Arkansas

James M. Walker, Department of Biological Sciences, University of Arkansas, Fayetteville, AR; jmwalker@uark.edu

Stanley E. Trauth, 13 Woodland Loop, Morrilton, AR; trauthse@outlook.com

We prepared this note to feature an example of an anuran species exhibiting an extended activity season and one that displays secretive and opportunistic behavior while inhabiting urban and suburban areas dominated by humans. For example, we have previously reported some of the activities of the Gray Treefrog (*Hyla versicolor* =

Dryophytes versicolor, Family Hylidae) on an urban residential lot in the city of Little Rock, Pulaski County, Arkansas (Walker et al. 2022). Herein, we stress fall activity of the species in two cities in Arkansas. In the northwest corner of the state of one of us (JMW) has lived on the same suburban residential property of 55 × 55 m for